

## Supplementary Data

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# The Transcriptomic Response of Mixed Neuron-Glial Cell Cultures to 1,25-Dihydroxyvitamin D3 Includes Genes Limiting the Progression of Neurodegenerative Diseases

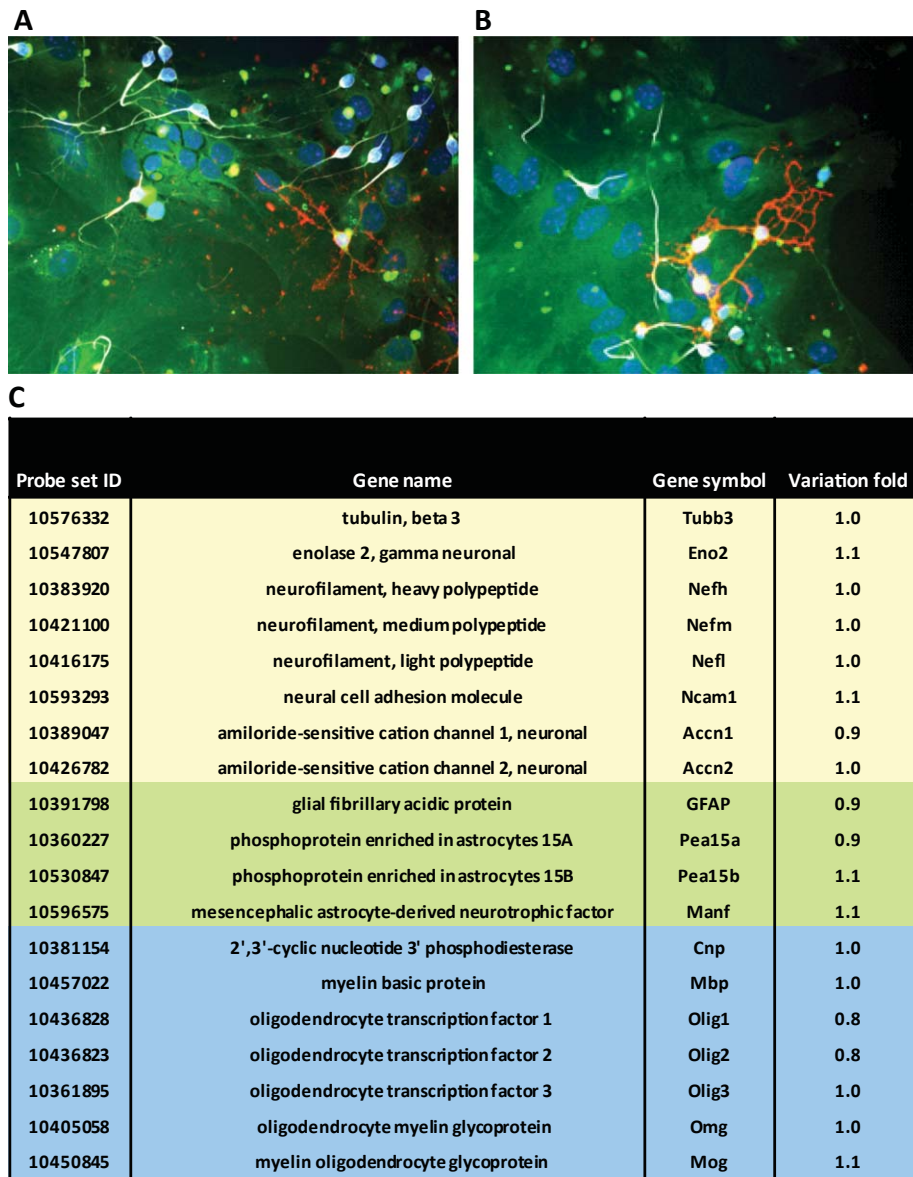
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Accepted 4 February 2013

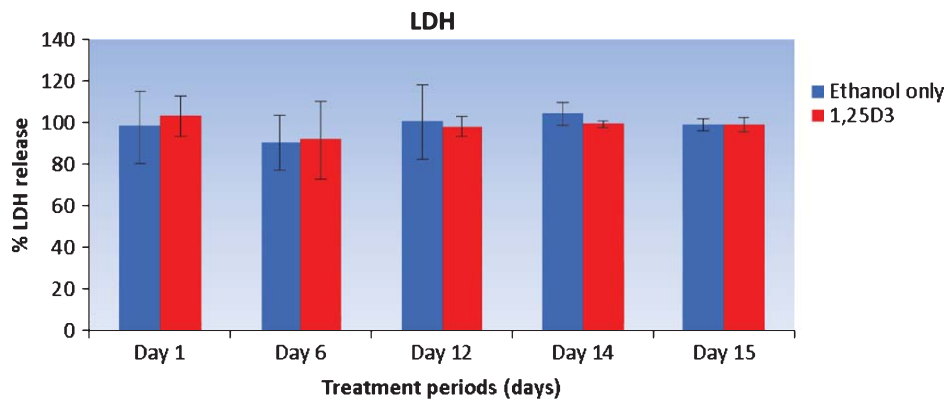
Supplementary Table 1  
Primers used for RT-qPCR experiments

Gene	Forward	Reverse
ACTB	ACCAGAGGCATACAGGGGACA	CTAAGGCCAACCGTGAAAAG
CAR14	GTTTCCCAGCCGCTTCTT	TGGGTCTAGGTGTGGGAATC
CBS	GCAGTGACAACCCCAAACA	GCCTGGTCAGGAGTGGTG
CRABP1	TGTGCAGTGAATCTTGTCTCA	AAGGTCGGAGAGGGCTTC
CYP24a1	TCTTGATTTGGGGGTGAAAA	CTGCCATTGCGTTCTGT
DIO2	GGAATTGGGAGCATCTTCAC	CTGCGCTGTGTCTGGAAC
FBLN1	CGGCACTGCTGCTTACAG	CAGGACCAGCTAAGATTCTT
GRPR	CTTCCGGGATTCGATCTG	TGATTTCAGAGTGCCTACAATCTTC
ITGA8	AGTTCTGTGCTCCTCTGGAA	TGGAGAATTCACATGGGGACT
ITIH3	CTCTGGGAGGCTCCGTTT	CTCTGGCTTGGAGACCTCTG
LCAT	GAGGGGGAGAAAACAAGTTGA	ACACGGCCTGTCATCCTC
NUPR1	GTGTGGTGTCTGTGGTCTGG	GAAGCTGCTGACCAAGTTCC
4833424O15Rik	GGAGCTAGGTTTCTCTGTAACCAC	AACCGTCCGATTTCTTGGGA
SLC7a3	GAGGAACAGCAGGCACCTT	AATTTCTGGGGTCATCTGGA
VDR	CTTCTCTGGGGACTCCTCTT	TGGACGAGTCCATCATGTCT
Tesc	AGGGTTTCCCGAGAGCAG	TCACGAAAGGAGAAGCTGAAAT
Cyp24a1	TCTTGATTTGGGGGTGAAAA	CTGCCATTGCGTTCTGT
SDHA	CAGTTCCACCCACAGGTA	TCTCCACGACACCCTTCTGT

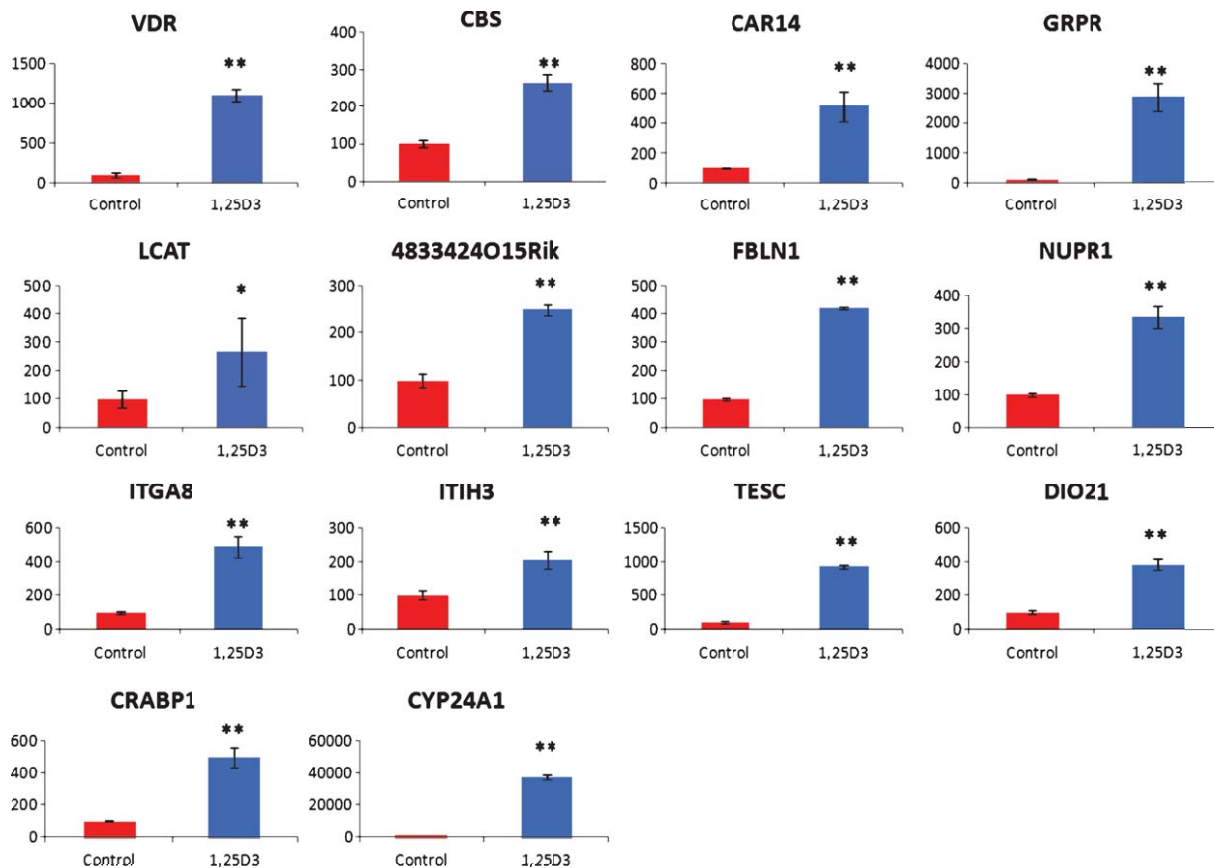
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Supplementary Figure 1. Immunophenotyping of mixed neuron-glia cell cultures. Mixed neuron-glia cell cultures prepared as described in the material and methods section were simultaneously stained with GFAP (astrocytes, green),  $\beta$ III-tubulin (neuron, white), and O4 (oligodendrocytes, red) antibodies. Nuclei were stained with DAPI (blue). A) Control culture (79.2% astrocytes (GFAP<sup>+</sup>), 18.2% neurons ( $\beta$ III-tubulin<sup>+</sup>), and 2.6% oligodendrocytes (O4<sup>+</sup>); number of counted cells = 1909. B) 1,25D3 treated culture (76.1% astrocytes (GFAP<sup>+</sup>), 21.4% neurons ( $\beta$ III-tubulin<sup>+</sup>), and 2.5% oligodendrocytes (O4<sup>+</sup>), number of counted cells = 1221). C) 1,25D3 treatment did not affect the expression of genes coding for specific markers for neurons (yellow), astrocytes (green), or oligodendrocytes (blue). Tubb3,  $\beta$ III-tubulin; Nef-H, neurofilament Heavy subunit; Nef-M, neurofilament medium subunit; Nef-L, neurofilament Light subunit; ENO2, enolase 2; Accn1, brain sodium channel 1; Accn2, brain sodium channel 2; Ncam1, neural cell adhesion molecule 1; GFAP, glial fibrillary acidic protein; Pea-15a, phosphoprotein enriched in astrocytes; Pea-15b, phosphoprotein enriched in astrocytes; Manf, mesencephalic astrocyte-derived neurotrophic factor; CNP, 2',3'-cyclic nucleotide 3' phosphodiesterase; MBP, myelin basic protein; Olig1, oligodendrocyte transcription factor 1; Olig2, oligodendrocyte transcription factor 2; Olig3, oligodendrocyte transcription factor 3; Omg, oligodendrocyte myelin glycoprotein; Mog, myelin oligodendrocyte glycoprotein



Supplementary Figure 2. Lactate dehydrogenase (LDH) release of mixed neuron-glia cells during the experiment. LDH release of the untreated group was considered 100%. The ethanol and 1,25D3 groups were not significantly changed compared to the untreated group ( $p > 0.2$  for all cases).



Supplementary Figure 3. Confirmatory RT-qPCR of the transcriptomic data for the genes newly identified as up-regulated by 1,25D3 and putatively involved in neurodegenerative or psychiatric diseases, or brain morphogenesis. Results are depicted relative to control and normalized to  $\beta$ -actin and succinate dehydrogenase complex, subunit A (SDHA) mRNA. (Student's *t* test \* $p < 0.05$ ; \*\* $p < 0.01$ ). VDR, vitamin D receptor; CBS, cystathionine-beta-synthase; CAR14, carbonic anhydrase XIV; GRPR, gastrin-releasing peptide receptor; LCAT, lecithin-cholesterol acyltransferase; FBLN1, RIKEN cDNA 4833424015, fibulin 1; NUPR1, nuclear protein, transcriptional regulator 1; ITGA8, integrin alpha 8; ITIH3, inter-alpha-trypsin inhibitor heavy chain 3; TESC, tescalcin; DIO2, deiodinase, iodothyronine, type II; CRABP1, cellular retinoic acid binding protein 1; Cyp24A1, 1,25-dihydroxyvitamin D3 24-hydroxylase.